www.nature.com/aja

RESEARCH HIGHLIGHT

A picture with more details is painted for prostate cancer

Wennuan Liu^{1,2}, Junjie Feng^{1,2} and Jianfeng Xu^{1,2,3}

Asian Journal of Andrology (2012) 14, 799-800; doi:10.1038/aja.2012.73; Published online: 30 July 2012

y comparing all gene-coding sequences in the genome between tumors and matched normal samples from clinically localized and castration-resistant (CR) prostate cancer patients, two groups of scientists have recently identified more than 5000 somatic mutations.^{1,2} These findings are important because they add another dimension of somatic DNA alterations in the genome of prostate tumors. Together with other known acquired DNA alterations in prostate tumors such as deletions, amplifications and fusions,³⁻⁶ they provide insights into the mechanisms of tumorigenesis and cancer progression of this heterogeneous disease. Some of the important findings are highlighted below.

SOMATIC MUTATION RATE IN PROSTATE TUMORS IS HIGHER THAN PREVIOUSLY ESTIMATED

It is commonly believed that point mutation is less prevalent in prostate tumors and was estimated at $0.33 \text{ Mb}^{-1.7}$ In these two studies, the average mutation rate was estimated at 1.4 Mb^{-1} in 112 untreated localized prostate tumors¹ and 2.0 Mb⁻¹ in 50 heavily treated castration-resistant (CR) tumors.² The higher mutation rate than previously estimated may reflect, at least in part, increased sequence coverage in these two studies. However, compared with other major human cancers, somatic mutation rate in prostate tumors remains generally low.⁷

COMMONLY AND SIGNIFICANTLY MUTATED GENES IN PROSTATE TUMORS

Although genes with recurrent mutations in prostate tumors have been documented (http://

Correspondence: Dr JF Xu (jxu@wfubmc.edu)

www.sanger.ac.uk/genetics/CGP/cosmic), they were based on candidate genes thought important in cancer development. Results from these two new reports, for the first time, present a comprehensive and objective list of the most commonly mutated genes in the genome of prostate tumors. Furthermore, each of these genes reached statistical significance, i.e., observed number of mutation is significantly higher than expected by chance given gene size, sequence context and frequency of mutations in each tumor. For localized prostate tumors, the 12 most significantly mutated genes are SPOP, FOXA1, TP53, PTEN, CDKN1B, MED12, THSD7B, SCN11A, NIPA2, PIK3CA, ZNF595 and C14orf49.1 SPOP was the most commonly mutated gene in these tumors, with a frequency of 13%. For CR tumors, the nine most significantly mutated genes include TP53, ZFHX3, RB1, PTEN, APC, AR, MLL2, OR5L1 and CDK12.² TP53 was at the top in this group, with a frequency of 40%.

COMMONALITY AND UNIQUENESS OF MUTATED GENES IN THESE TWO TYPES OF TUMORS

PTEN and TP53 were the only genes significantly mutated in both localized and CR tumors in these two studies, emphasizing their broad roles in cancer initiation, progression and treatment resistance. For majority of the remaining genes, recurrent mutations were also found in both types of tumors, although statistical significance was reached in only one type of these tumors. There are, however, several exceptions. PIK3CA, ZNF595 and C14orf49 were significantly mutated in localized prostate tumors but no mutation in these genes was observed in CR cancer, suggesting that mutations of these genes unlikely play an important role in the development of lethal CR prostate cancer. On the other hand, of the nine significantly mutated genes in CR tumors, mutation in three genes (AR, RB1 and CDK12) was not found in localized prostate tumors. Mutations of these genes in CR tumors may be triggered in response to hormone therapy, or reflect selection advantage for lethal CR prostate cancer.

TWO TYPES OF MUTUALLY EXCLUSIVE PROSTATE TUMORS

By examining tumors with SPOP mutations, Barbieri et al.1 found that none of these tumors carried ETS family gene rearrangement. Furthermore, they found that tumors with SPOP mutations were positively associated with deletion of CHD1 at 5q21.1 and deletions of FOXO3 and PRDM1 at 6q21. Based on these observations, Barbieri et al.1 proposed that SPOP mutations may define a new molecular subtype of prostate cancer. Similarly, using an integrated analysis of exome sequencing, copy number and expression, Grasso et al.² found an inverse association between ETS family gene rearrangement and focal deletion or mutation of CHD1. Therefore, both studies suggest two distinct and mutually exclusive prostate cancers: (i) tumors with ETS family gene rearrangement, and (ii) tumors with deletions or mutations at CHD1/SPOP. It is, however, unclear whether CHD1 or SPOP defines the second subtype of prostate cancer. It is noted that deletion of CHD1 was previously reported to be inversely associated with the genomic deletion that resulted in TMPRESS-ERG in localized prostate tumors.8

FOXA1 AND CHROMATIN/HISTONE-MODIFYING GENES PHYSICALLY INTERACT WITH AND FUNCTIONALLY REGULATE AR

In the mutational landscape of CR prostate cancer identified by Grasso *et al.*, it is noteworthy that multiple recurrently mutated

¹Center for Cancer Genomics, Wake Forest University School of Medicine, Winston-Salem, NC 27157, USA; ²Center for Human Genomics and Personalized Medicine Research, Wake Forest University School of Medicine, Winston-Salem, NC 27157, USA and ³Fudan Institute of Urology, Fudan University, Shanghai 200040, China

genes are commonly involved in the modulation of androgen-AR signaling.² Besides alterations that are directly associated with deregulated androgen synthesis (e.g. CYP11B1 amplification) and signaling (e.g. AR amplification/mutation, point mutations of NKX3-1), recurrent copy number and mutational alterations were also identified in genes whose protein products have been demonstrated to physically interact with and functionally regulate AR. Among these genes include the AR collaborating factor FOXA1, whose mutations were found in 3.4% of prostate cancer patients and whose mutant forms were shown to repress androgen signaling and increase tumor growth. In addition, recurrent mutations were notably found in multiple chromatin/histone-modifying genes, including MLL2 (mutated in 8.6% of prostate cancer), ASXL1 and UTX, which also interact with and regulate AR signaling. These findings together suggest that the aberrant androgen-AR signaling caused by alterations of these recurrently mutated genes may serve as at least one common mechanism underlying the castration resistance phenotype exhibited by almost all prostate cancer patients undergoing hormonal therapy.

ETS2 IS A CANDIDATE TUMOR-SUPPRESSOR GENE

An important finding in the study of Grasso et al. is somatic alterations at ETS2 and their role in prostate cancer development and invasion. ETS2, located between TMPRESS2 and ERG, is deleted in one-third of prostate cancers and mutated in a CR tumor. Its tumor-suppressor role was also suggested from the observation that tumors with TMPRESS-ERG fusions through deletion were more aggressive than those through translocation. More importantly, the tumorsuppressing function of ETS2 was suggested by the demonstration that overexpression of wild-type ETS2 led to decreased cell proliferation, migration and invasion, and that mutant ETS2 had opposite effects.

Future studies should be extended to intergenic/noncoding regions in the genome to better define chromosomal rearrangements, including types, boundary, and frequency of deletions, gains and fusions. These types of alterations are important in prostate tumors but were only examined using low-resolution methods such as array-comparative genomic hybridization and genome-wide singlenucleotide polymorphism arrays or in a few samples using whole-genome analysis.³ In addition, greater efforts should be devoted to assessing correlation of somatic alterations with clinical presentations.

- Barbieri CE, Baca SC, Lawrence MS, Demichelis F, Blattner M et al. Exome sequencing identifies recurrent SPOP, FOXA1 and MED12 mutations in prostate cancer. Nat Genet 2012; 44: 685–9.
- 2 Grasso CS, Wu Y-M, Robinson DR, Cao X, Dhanasekaran SM et al. The mutational landscape of lethal castration-resistant prostate cancer. *Nature*; e-pub ahead of print 20 May 2012; doi: 10.1038/ nature11125.
- 3 Berger MF, Lawrence MS, Demichelis F, Drier Y, Cibulskis K et al. The genomic complexity of primary human prostate cancer. Nature 2011: 470: 214–20.
- 4 Liu W, Laitinen S, Khan S, Vihinen M, Kowalski J et al. Copy number analysis indicates monoclonal origin of lethal metastatic prostate cancer. Nat Med 2009; 15: 559–65.
- 5 Taylor BS, Schultz N, Hieronymus H, Gopalan A, Xiao Y et al. Integrative genomic profiling of human prostate cancer. Cancer Cell 2010; 18: 11–22.
- 6 Tomlins SA, Rhodes DR, Perner S, Dhanasekaran SM, Mehra R et al. Recurrent fusion of TMPRSS2 and ETS transcription factor genes in prostate cancer. *Science* 2005; **310**: 644–8.
- 7 Kan Z, Jaiswal BS, Stinson J, Janakiraman V, Bhatt D et al. Diverse somatic mutation patterns and pathway alterations in human cancers. *Nature* 2010; **466**: 869–73.
- 8 Liu W, Lindberg J, Sui G, Luo J, Egevad L et al. Identification of novel CHD1-associated collaborative alterations of genomic structure and functional assessment of CHD1 in prostate cancer. Oncogene; epub ahead of print 5 December 2012; doi: 10.1038/ onc.2011.554.

800